

Maternal undernutrition and endocrine development

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Jane E Harding[†],
José GB Derraik and
Frank H Bloomfield

[†]Author for correspondence
Liggins Institute, University of
Auckland, Private Bag 92019,
Auckland, New Zealand
Tel.: +64 9373 7599 ext 86439
Fax: +64 9373 7497
j.harding@auckland.ac.nz

Maternal undernutrition, whether it occurs before conception, throughout gestation or during lactation, may lead to physiological adaptations in the fetus that will affect the health of the offspring in adult life. The timing, severity, duration and nature of the maternal nutritional insult may affect the offspring differently. Other factors determining outcome following maternal undernutrition are fetal number and gender. Importantly, effects of maternal undernutrition may be carried over into subsequent generations. This review examines the endocrine pathways disrupted by maternal undernutrition that affect the long-term postnatal health of the offspring. Maternal and childhood undernutrition are highly prevalent in low- and middle-income countries, and, in developed countries, unintentional undernutrition may arise from maternal dieting. It is, therefore, important that we better understand the mechanisms driving the long-term effects of maternal undernutrition, as well as identifying treatments to ameliorate the associated mortality and morbidity.

KEYWORDS: developmental origins of health and disease • fetal growth • fetus • glucose–insulin axis • hypothalamic–pituitary–adrenal axis • maternal undernutrition

There is accumulating evidence that events in early life confer risks for the onset of adult diseases: the so-called developmental origins of health and disease [1,2]. Environmental influences may lead an organism along certain pathways during sensitive, and often brief, periods of development, so that it may develop characteristics better adapted to the environment in which it is likely to live [3]. This developmental plasticity during intrauterine development is beneficial if the extrauterine environment is consistent with that signaled via the mother before birth. However, if the extrauterine environment turns out to be very different, plasticity may also result in adaptations *in utero* that are inappropriate for the environment in adulthood and, therefore, detrimental to health.

Any environmental effect on fetal or postnatal development must ultimately act via altered gene expression. Epigenetic regulation of gene transcription is likely to be a key mechanism through which fetal nutrient supply can alter gene expression in the developing fetus in response to environmental cues. Gene expression may be modified by a number of processes, primarily DNA methylation and histone modification [4], both of which may be altered by the availability of amino acids and micronutrients during development *in utero* [5,6]. For example, folic acid deficiency is associated with reduced DNA methylation in

humans [7]. Similarly, feeding genetically identical mice a diet high in methyl donors, including folate, results in epigenetic modification of gene expression in the offspring [8]. Importantly, these modified characteristics are transmitted to subsequent generations, demonstrating that epigenetic mechanisms can result in heritable traits that may reflect persistent adaptations to environmental cues over several generations [9]. Thus, DNA methylation-based markers, for example, are seen as promising tools for clinical diagnostics and therapeutics [10], and screening for epigenetic markers in early life may lead to individually customized interventions to reduce the risk of later disease [11].

The intrauterine environment is affected particularly by both overabundance and deficiency of fetal nutrient supply. The proportion of the population in the developed world who are overweight and obese is steadily increasing. In women of childbearing age, this may result in a dietary intake during pregnancy that is high in calories and rich in fat and carbohydrates [12,13]. This can then lead to intrauterine fetal overnutrition, which is associated with numerous deleterious effects for the offspring [14], ranging from higher prevalence of congenital anomalies [15,16] to obesity, metabolic syndrome, and Type 2 diabetes in adult life [12,17–20].

On the other hand, it has generally been assumed that, except under severe conditions, maternal nutrition is not a limiting factor for fetal development and growth. However, deficiencies in the maternal diet can also have significant effects on the fetus, permanently affecting health in later life [21]. For example, fetal adaptations to intrauterine undernutrition, likely resulting from inadequate materno-placental nutrient supply, increases the risk of cardiovascular diseases in adulthood [22]. Similarly, there are extensive data on the long-term effects of severe food restriction in human pregnancy, resulting from exposure to the Dutch famine of 1944–1945 [23].

Undernutrition may affect the offspring differently, depending on its timing, severity, duration and the nature of the insult. Other factors altering the effects of maternal undernutrition are the number of fetuses in a given pregnancy and fetal gender. Furthermore, although some effects may result directly from alterations in substrate availability, many are thought to be mediated by hormonal effects [22]. In this review, we focus primarily on the endocrine pathways that are disrupted by maternal undernutrition and the consequent long-term effects on postnatal health of the offspring.

Factors modifying the effects of maternal undernutrition on fetal endocrine development

Nature of undernutrition

There is evidence that altered maternal macronutrient balance [24], micronutrient intake [25] and overall caloric intake [23] can each influence postnatal disease risk. Although the majority of studies of the effects of maternal undernutrition on the developing fetus have investigated global undernutrition, many have examined the effects of deficiencies of particular dietary components.

Micronutrient deficiency in human pregnancy has been associated with a number of adverse outcomes, such as preterm birth, intrauterine growth restriction (IUGR) and congenital defects [21]. The importance of an adequate micronutrient supply in pregnancy has been supported by animal experiments, as well as the study of clinical conditions in humans associated with severe deficiency of particular vitamins and minerals [21]. In rats, a maternal vitamin-restricted diet (50% of normal intake of vitamins A, B₁, B₂, B₃, B₅, B₆, B₉, B₁₂, D₃, E, H and K₁) from preconception to weaning led to increased body fat content and plasma triglycerides, as well as lower lean body mass in adult offspring [26]. However, most of these effects appear to be reversed with postinsult vitamin supplementation [26].

Mild iron deficiency during pregnancy in rats led to offspring with significantly increased blood pressure in adult life [27]. Iron deficiency is the most common cause of nutritional anemia in humans, and in pregnancy it leads to elevated IGF-1 and reduced thyroxine concentrations in cord blood [28]. In addition, the more severe the anemia, the greater the effects on fetal growth, which probably result from the numerous effects of anemia on endocrine systems, including the somatotrophic, thyroid and hypothalamic-pituitary-adrenal (HPA) axes [28].

Iodine is an essential component of the two thyroid hormones triiodothyronine (T₃) and thyroxine (T₄) [29]. In humans, iodine deficiency during pregnancy leads to hypothyroidism, which,

when sustained during a critical window of development from conception to 2 years of age, hinders brain development, causing irreversible mental retardation [29].

Dietary zinc restriction in rats during pregnancy and lactation induces functional and morphological adaptations in the fetuses that result in increased blood pressure in adult offspring [30]. Renal effects include reduced glomerular number, glomerular and tubular fibrosis, increased apoptosis and reduced antioxidant activity [30]. Increased reactive oxygen species may also contribute to the apoptosis and renal damage associated with zinc deficiency [30].

A calcium-deficient diet in rats is also associated with hypertension in adult offspring [31]. One randomized controlled trial in humans showed that calcium supplementation during pregnancy is associated with lower systolic blood pressure in the offspring at age 5–9 years, particularly among overweight children [32]. However, an Australian study indicated that the potential long-term benefits of gestational calcium supplementation on offspring blood pressure may be apparent only in the offspring of mothers with high blood pressure during pregnancy [33]. The associated endocrine mechanisms are still unclear.

Protein deficiency during pregnancy has been frequently investigated. Early studies showed an inverse relationship between maternal protein intake and the systolic blood pressure of rat offspring, and that the hypertension in offspring from low-protein-fed dams (9 vs 18% protein) was associated with increased pulmonary angiotensin-converting enzyme activity [34]. Other effects on the cardiovascular system include decreased aortic wall thickness and elastin content [35], as well as structural and functional cardiac changes, predisposing the animals to impairment of diastolic and systolic function in later life [36]. Other studies have consistently demonstrated hypertension in offspring following maternal protein restriction, with some also reporting increased fetal mortality [37]. In humans, an isocaloric low-protein diet in pregnancy has also been associated with increased systolic blood pressure in adolescent boys, independent of birthweight and maternal triceps skinfold thickness during pregnancy [38]. However, one prospective human study showed no association of low protein intake during pregnancy with offspring blood pressure at 6 months [39].

Low-protein diets throughout gestation also have been shown to decrease serum estradiol concentrations in female offspring [37], impair estrous cyclicity [40], alter the offspring's fat distribution and food intake in adulthood [41], and alter glucose tolerance of young rats [42]. Furthermore, the male offspring of rat dams on a protein-restricted diet during pregnancy had elevated cholesterol and triglyceride concentrations, and showed insulin resistance as adults, while the body composition of females had increased relative levels of tissue fat and decreased protein [43]. In mice, a low-protein diet during pregnancy predisposed genetically susceptible mice to the development of atherosclerosis [44].

Note that the deleterious effects on the offspring of a protein-deficient diet in pregnancy are magnified by diet in postnatal life. The offspring of rat dams subjected to protein undernutrition throughout gestation and lactation become significantly more obese, insulin resistant, hyperlipidemic and hypertensive, as well

as more sedentary as adults, when fed a highly palatable diet from weaning [19,45,46]. Furthermore, protein malnutrition throughout gestation shortened the lifespan of mice when dams were allowed to overfeed during lactation [47].

Timing of undernutrition

Retrospective studies on offspring of women exposed to the Dutch famine have shown that early gestational undernutrition primarily affects the cardiovascular system, leading to a greater incidence of coronary heart disease, a more atherogenic lipid profile and disturbed blood coagulation profiles [23]. Furthermore, the offspring of mothers exposed to famine in early gestation had increased stress responsiveness, a greater incidence of obesity, impaired glucose tolerance, and female offspring were more likely to develop breast cancer [23]. The increased incidence of obesity in adults whose mothers were exposed to the Dutch famine during early pregnancy [23,48], in particular, may be a direct consequence of adaptations in the endocrine sensitivity of fetal adipose tissue [49].

Animal experiments have shown that maternal nutrition before and in very early pregnancy can have important effects on the health of the offspring. In rats, undernutrition of dams for 8 weeks before conception affects growth of vital organs and results in increased blood glucose and cholesterol concentrations in the adult offspring [50]. The induction of hypertension in the offspring was also more marked when the low-protein maternal diet started very early in gestation compared with when it started in mid gestation [37]. Indeed, maternal protein undernutrition for just the first 4.5 days (blastocyst stage) of rat pregnancy (duration: 21 days) results in postnatal hypertension of the offspring [51]. Embryo transfer experiments suggest that the effects are intrinsic to changes in the blastocyst, rather than the intrauterine environment [52].

In sheep, where it is possible to study fetal growth and endocrine function *in vivo*, it is now abundantly clear that maternal undernutrition in the periconceptual period alters many aspects of fetal development in late gestation. Fetal growth trajectory is reduced in late gestation following periconceptual undernutrition [53]. However, fetuses of ewes undernourished only before conception show greater fetal growth and metabolic responses to an acute maternal fast in late gestation than fetuses of ewes undernourished both before and after conception [54], with few changes seen in fetuses of ewes undernourished only after conception. These differences cannot be explained by changes in the fetal glucose–insulin axis or the fetal or maternal HPA axis at the time of the fast [54]. Thus, it appears that maternal nutrition before conception is particularly important in determining fetal responses to late gestational stress, and that effects of maternal undernutrition on fetal development are due neither solely to limitation of substrates for tissue accretion nor to excess fetal glucocorticoid exposure at the time of undernutrition [54].

Periconceptual undernutrition also alters several aspects of endocrine regulation in late-gestation fetal sheep [53], and appears to accelerate maturation of the insulin and HPA axes [55,56]. Early gestation nutrient restriction promotes adipose tissue deposition in

the offspring [49]. Offspring of ewes undernourished in the periconceptual period have impaired glucose tolerance, which worsens with increasing age [57]. Furthermore, maternal periconceptual undernutrition alters behavioral laterality in offspring in a sexually dimorphic manner [58].

Maternal exposure to the Dutch famine in mid gestation was associated with increased incidence of microalbuminuria and obstructive airways disease, as well as impaired glucose tolerance, in the adult offspring [23]. In sheep, mid-gestation undernutrition results in increased adipose tissue deposition [49] and increased expression of genes that regulate apoptosis in the fetal ovary [59], as well as reduction of large corpora lutea [60].

Exposure to the Dutch famine in late gestation tended to affect intermediary metabolism in the offspring, particularly glucose–insulin homeostasis, leading to an increased risk of Type 2 diabetes [61,62]. Similarly, glucose tolerance was reduced only in the 1-year-old offspring of ewes undernourished in late gestation but not of ewes undernourished in early gestation [63]. Brief, profound, maternal undernutrition in late gestation in sheep also increased central HPA axis responses to corticotrophic stimulation in adult offspring, despite the fact that birthweight was not affected [64]. Interestingly, if the period of undernutrition was prolonged to an extent that did reduce birthweight, the effects on HPA axis function were mitigated, confirming that reduced size at birth following maternal undernutrition is not required for altered postnatal physiology [64].

Fetal number & gender

The effects of maternal undernutrition on the offspring are also affected by fetal number [49,65]. For example, nutrient restriction causes an upregulation of specific mitochondrial proteins within the adipose tissue of twin fetuses but not of singletons [49]. One study found that, in most cases (depending on timing of undernutrition), mRNA expression for *IGF-1* and *-2*, their receptors, the growth hormone (GH) receptor and the glucocorticoid receptor, were all significantly downregulated in the kidneys of twin fetuses but not in those of singletons [66]. Periconceptual undernutrition also results in an earlier prepartum activation of the central HPA axis in twin fetal sheep compared with singleton fetuses [67], but delayed maturation of the adrenal gland was already present in twin fetuses in early gestation, perhaps to prevent preterm onset of parturition [68]. Periconceptual undernutrition also abolished the accelerated maturation of the glucose–insulin axis that is seen in twin fetal sheep compared with singletons [69].

The interactions between maternal nutrition and fetal number persist into postnatal life, with the effects of undernutrition on behavioral laterality being different in singletons and twins [58]. These data suggest that the physiology of twin pregnancies is quite different from that of singletons, and likely to be determined by a combination of factors acting in both early and late gestation [69,70]. The late gestation effects may represent an additional nutritional challenge to twin fetuses compared with singletons, but the effects of fetal number on aspects of development are not the same as early pregnancy undernutrition, suggesting that the early pregnancy effects may be mediated by different mechanisms.

Fetal sex also plays a significant role in determining the outcomes associated with maternal undernutrition. In humans, a low-protein diet during pregnancy was associated with increased systolic blood pressure in adolescent boys but not girls [38]. A low-energy diet in the initial two-thirds of pregnancy in rats produced male offspring that spontaneously increased their intakes of a low-fat diet, gained more weight beginning at weaning and developed diet-induced obesity as adults [71,72]; these effects were not observed in females. Another study in rats observed that gestational undernutrition produced male offspring with reduced bodyweight, while females had increased bodyweight and adiposity as adults [73]. The effects of gestational and postnatal protein restriction on renal angiotensin receptor expression were also sex specific in rats [74] and mice [75], as were the effects of protein restriction during lactation on plasma leptin concentrations in rats [43].

In guinea pigs, acute maternal nutrient restriction for 48 h during the period of maximal fetal brain growth resulted in reduced basal cortisol and adrenocorticotrophic hormone (ACTH) in adult male offspring but led to elevated basal cortisol and normal ACTH in female offspring [76]. Indeed, in young adult lambs, early-gestation undernutrition seemed to have a minor influence on HPA axis function when considered alongside the effect of sex *per se* [77]. These observed differences are dependent upon sex hormones and/or sexually dimorphic development of central pathways regulating energy homeostasis [19]. Similarly, the onset of hypertension in rats following a low-protein diet was delayed in females [37], most likely a result of the effects of estrogen [78]. Epigenetic modification may underlie some of these sex-specific effects, particularly as both the promoters of androgen and estrogen receptor genes, and the expression of their target genes, have been shown to be regulated by epigenetic mechanisms [79].

Effects on endocrine development

Renin–angiotensin system

A well-recognized long-term effect of maternal undernutrition in the offspring is increased propensity to hypertension, which probably results from the interaction of a number of endocrine systems. However, there is accumulating evidence that the effects of maternal undernutrition on nephrogenesis and the renin–angiotensin system (RAS) play a major role.

Maternal food restriction alters gene expression of fetal renal transcription factors and growth factors in rats, impairing nephrogenesis during fetal development [80,81]. Maternal undernutrition in mice [75], rats [81–84] and sheep [85] reduces nephron numbers in the offspring. In humans, a Dutch famine cohort study linked maternal starvation to impaired renal function in the offspring; more specifically, a mid-gestation insult led to higher rates of microalbuminuria, which was likely associated with reduced glomerular endowment [86].

Undernutrition of rats in late gestation did not affect glomerular ultrastructure in the offspring, but rather resulted in reduced nephron number, decreased glomerular filtration rate and increased blood pressure [87]. Other effects of maternal undernutrition include the inhibition of vascular EGF expression in

microvascular and aortic endothelial cells early in life, resulting in decreased angiogenesis and increased peripheral vascular resistance, both of which may contribute to offspring hypertension [88]. In experimental animals, the reduction in nephron numbers alone is associated with hypertension in later life [66,84,85], and analogous observations have been made in human studies [89].

However, a reduction in nephron number likely interacts with alterations in the RAS leading to a detrimental outcome. In rats, maternal protein restriction during pregnancy suppressed the newborn intrarenal RAS, with reduced renin mRNA expression in the kidney, as well as suppressed angiotensin II concentrations in renal tissue [90]. Similarly, abnormal kidney RAS ontogeny was observed in the offspring of protein-restricted rat dams, associated with neonatal suppression of angiotensin receptor 1 protein abundance in the newborn kidney but upregulation at 4 weeks of age [91]. However, receptor mRNA levels and sensitivity were upregulated as these animals aged [91–93], potentially resulting in inappropriate reductions in glomerular filtration rate. In sheep, maternal undernutrition also results in reduced nephron endowment and altered renal protein levels of angiotensin II converting enzyme and the type 2 angiotensin receptor in young adult offspring [85]. Intriguingly, treatment of the offspring of undernourished rats with a specific angiotensin II receptor antagonist in early postnatal life prevents the later development of increased blood pressure, providing further evidence that angiotensin II may play a major role in adult hypertension following maternal protein undernutrition during pregnancy [94].

Adipokine axis

The greater incidence of obesity in the adult offspring of mothers exposed to famine during gestation [95] may result from adaptations in the endocrine sensitivity of fetal adipose tissue [49]. Maternal undernutrition during early-to-mid gestation in sheep, for example, led to increased bodyweight and fat deposition in adolescence, as well as dysregulated glucose uptake, despite the absence of any change in birthweight [96].

An undernutrition insult over the period of maximal placental growth in sheep (early-to-mid gestation) resulted in lower maternal plasma cortisol, leptin, T_4 and IGF-1 concentrations, accompanied by increased fetal adipose tissue deposition and an upregulation of mRNA levels of the IGF receptors in this tissue [97]. The offspring of ewes similarly undernourished had increased adiposity at term with upregulation of uncoupling protein 2 (*UCP2*) and *PPAR- α* mRNA levels in adipose tissue [98,99]. It was, therefore, suggested that a combination of raised *PPAR- α* and *UCP2*, in conjunction with increased sensitivity to IGFs owing to upregulation of their receptors (but not their ligands) within the adipocyte, could explain the greater fat mass in the fetuses of undernourished mothers [97]. In addition, enhanced *UCP2* abundance could enhance apoptosis rate [100] and compromise adipose tissue function during periods of environmental stress [99], increasing adult predisposition to disease [49].

Nonetheless, most fetal adipose tissue is accrued in the final trimester [101]; over this period there is an increased abundance within the fetal circulation of hormones that are important in

regulating fetal adipose tissue growth [49,102], including IGF-1 and leptin [103,104]. Maternal leptin crosses the blood–placenta barrier into the fetal circulation [105], but the female offspring of rats fed a protein-restricted diet solely during lactation also had mean serum leptin concentrations that were nearly half those in controls [43], suggesting that the effects are on the adipokine axis of the offspring rather than the mother. Studies of genetically manipulated mice suggest that the presence of leptin and associated energy regulation are integral to the acceleration of obesity when on a high-fat diet following undernutrition *in utero*, and the premature leptin surge likely plays an essential role as a signal during the early neonatal period in the developmental origins of obesity [106].

It seems that perinatal perturbations of leptin action, either through reduced leptin availability or through hypothalamic leptin resistance, is a mechanism by which perinatal undernutrition may permanently alter hypothalamic circuits [107]. Maternal undernutrition from conception to the end of the period of maximal placental growth in sheep led to an increased leptin response to norepinephrine and angiotensin II infusions [108]. Undernutrition between 30 and 80 days of gestation, a period during which early fetal brain development takes place in sheep, resulted in reduced expression of neuropeptide Y (*NPY*) in the hypothalamus in 1-week-old offspring [109]. If these offspring were placed on an obesogenic diet after weaning, the prenatal effect of maternal undernutrition amplified the effect of postnatal obesity, with obese offspring of undernourished ewes having significantly higher leptin, insulin and nonesterified fatty acid concentrations compared with obese offspring of well-nourished ewes [109]. These changes were accompanied by increased hypothalamic mRNA levels of the melanocortin-4 receptor, a key part of the hypothalamic appetite regulatory pathway. The authors interpreted these changes as reflecting an altered adaptation in energy balance when juvenile obesity follows upon prenatal undernutrition [109].

In rats, the offspring of undernourished dams had premature onset of neonatal leptin surge, impaired leptin transport to the brain, increased density of hypothalamic nerve terminals and elevation of endogenous serum leptin concentrations [110]. The premature leptin surge was associated with adult obesity, perhaps by altering energy regulation by the hypothalamus [110]. Maternal undernutrition in mid-to-late gestation in rats dramatically reduced postnatal plasma leptin concentrations and hypothalamic mRNA levels of the leptin receptor in male pups [111], and affected development of pro-opiomelanocortin (POMC) neurons in the arcuate nucleus [112]. Leptin has also been shown to promote neuronal outgrowth from the arcuate nucleus to the paraventricular nucleus of the developing hypothalamus during the lactation period in rats, thus, potentially ‘hardwiring’ the hypothalamic appetite regulatory system [18]. Therefore, the evidence from rats shows that maternal undernutrition leads to alterations in the hypothalamus of adult offspring through its effects on leptin pathways, differently programming the long-term appetite regulatory system, especially the responses of POMC neurons to energy status and food-intake rhythm [111]. Interestingly, neonatal leptin treatment in female rats permanently reverses

the long-term effects of maternal undernutrition, normalizing caloric intake, locomotor activity, bodyweight, fat mass, as well as fasting plasma glucose, insulin and leptin concentrations in adult life [113]. The authors attributed this effect, at least partly, to the resetting of central and/or peripheral pathways that regulate energy homeostasis [113].

Gonadotrophic axis

There is considerable evidence that reduced maternal nutrient intake during pregnancy can affect reproductive function in the adult offspring [114]. Furthermore, the effects of maternal undernutrition have also been demonstrated on the reproductive tracts of fetuses [114], suggesting one possible explanation for the transmission of effects to the second generation that have been reported in humans [115] and rats [116,117]. Maternal undernutrition in early gestation in sheep led to increased expression of steroidogenic acute regulatory protein (*StAR*) mRNA in the fetal testes, and increased plasma testosterone concentrations [118]. In female fetuses, ovarian development is affected [116,117], with altered proliferation and expression of apoptosis-regulating genes [59] and increased numbers of small follicles in the ovary [60]. There are also effects on the central gonadotrophic axis, with increased pituitary sensitivity to GnRH [60].

Maternal undernutrition affects the gonadotrophic axis of the offspring not only when the insult is restricted to the gestational period alone, but also when it occurs solely during lactation. In rats for example, maternal protein and energy, or solely energy, restriction during lactation lowered serum estradiol and raised testicular testosterone concentrations, and also enhanced androgen receptor expression whilst lowering estrogen receptor and aromatase mRNA expression levels in the testis of offspring at weaning [119]. Protein and energy restriction in lactation also led to significantly elevated concentrations of serum testosterone [119].

The onset of sexual maturity in the offspring is also affected by maternal undernutrition. Little evidence exists for humans, but it has been reported that girls of lower birthweight have an earlier onset of menarche; this effect is enhanced by obesity in childhood [120]. Furthermore, women exposed to the Dutch famine *in utero* delivered their first child at a younger age than those not exposed [121]. In sheep, moderate maternal undernutrition was not detrimental to the onset of puberty (defined as first ovulation) in female lambs, but it delayed the onset and magnitude of sexual activation in male offspring, and lowered peak testosterone concentrations [122]. In rats, late-gestational undernutrition resulted in delayed onset of puberty in both male and female offspring [123]. A more recent study showed that maternal protein restriction solely during lactation also delayed onset of puberty in female rats, associated with decreased luteinizing hormone (LH) and increased testosterone serum concentrations at 70 days, with the latter difference persisting until 1 year of age [124]. Significant reduction in serum LH at 70 days was also observed in rats exposed to undernutrition only during pregnancy [124].

In a recent study in rats, maternal undernutrition during pregnancy led to early onset of puberty, as well as to a reduction in proestrus progesterone levels in later life [125]. The reasons for

this contrasting result are unclear but it appears that, in rats, an undernutrition insult, restricted to or including the lactation period, delays sexual maturation, while undernutrition solely during gestation may have an opposite effect. The severity of the undernutrition insult may also affect the outcome.

Furthermore, maternal undernutrition not only affects the timing of sexual maturation but may also affect reproductive capacity. An earlier study showed that maternal undernutrition during gestation and lactation in rats significantly impaired ejaculatory capacity in male offspring, as well as reducing absolute weight of testicles, and absolute and relative weights of seminal vesicles [126]. However, data from the Dutch famine suggested that human *in utero* exposure to severe food limitation improved rather than impaired reproductive success in women (but not men), with exposed women giving birth to a larger number of children and being less likely to be childless [121]. Interestingly, the rate of twinning was approximately three-times that of unexposed women, suggesting that the increased number of offspring may, in part, reflect alterations in the regulation of ovulation or other aspects of reproductive function [121].

Glucose–insulin axis

In humans, prenatal exposure to famine at any time, but especially during late gestation, is associated with decreased glucose tolerance in adult life [62,127]. Small-for-gestation-age (SGA) babies or those with IUGR are often chronically starved *in utero* [128–130], and, thus, provide some evidence regarding the later effects of intrauterine undernutrition. Short prepubertal IUGR children have been shown to have impaired insulin sensitivity in comparison to their normal-birthweight peers [131]. Young adults born with IUGR also had elevated plasma concentrations of insulin and proinsulin, indicating early development of insulin resistance [132]. Similarly, 25-year-old people born with IUGR displayed decreased insulin-stimulated glucose uptake without major impairment of insulin secretion [133].

In sheep, maternal periconceptional undernutrition appears to accelerate fetal pancreatic maturation [56,69]. However, the postnatal offspring of undernourished ewes demonstrated decreasing glucose tolerance with increasing postnatal age [57], suggesting that the *in utero* adaptation may have resulted in maturation at the expense of differentiation. Late-gestation undernutrition in sheep during the period of maximal fetal growth also results in impaired glucose tolerance and insulin resistance in the adult offspring [63], as does late-gestation maternal undernutrition in rats [46,134].

The mechanisms underlying the impaired glucose tolerance in offspring of undernourished mothers have not been entirely elucidated, but there may be effects at several levels. These include reduced β -cell proliferation, reduced islet size and vascularization in the fetal pancreas [135], as well as impaired β -cell neogenesis resulting in a reduced pancreatic β -cell mass and insulin content [136,137]. In addition, signaling pathways within the β -cell may be altered [138], and there is evidence that in adult offspring, β -cell telomere length is shortened, indicating greater cell senescence [139]. Peripheral organ responses may also play a role, with altered glucose uptake in adipose tissue, mediated via reduced

expression of the glucose transporter GLUT4 [63], and insulin resistance in adipose tissue and muscle [46]. Recently, studies in rats with growth restriction induced by uterine artery ligation have demonstrated that a key pancreatic transcription factor is downregulated by epigenetic mechanisms [140]. Whether the same is true following maternal undernutrition is not yet clear.

Importantly, the effects of maternal undernutrition may be carried over into subsequent generations [19]. For example, when the mothers of an F1 generation were subjected to a low-protein diet, insulin resistance was observed in second-generation rats [116,141]. The female offspring of undernourished rats displayed decreased β -cell mass and were later unable to adapt their endocrine pancreatic mass to pregnancy, while the subsequent F2 generation displayed a reduction in β -cell mass and numerical density of islets [142].

Hypothalamic–pituitary–adrenal axis

Glucocorticoids have been proposed as key factors mediating the long-term effects of an adverse intrauterine environment [143], as they can alter important gene transcription in the developing fetus, trigger cell differentiation in many tissues *in utero*, stimulate the switch from growth to maturation [144–146] and affect the secretion of a number of metabolic hormones [138,147]. An adverse intrauterine environment, such as that resulting from maternal undernutrition or prenatal maternal stress, may result in increased maternal and, thus fetal circulating concentrations of glucocorticoids [143,148,149].

However, although an acute maternal undernutrition insult may elevate circulating glucocorticoid concentrations [148,149], it is less clear that chronic undernutrition has a similar effect. Indeed, in sheep undernourished for several weeks, maternal basal and stimulated glucocorticoid concentrations were actually decreased [97,150]. An additional factor determining the effects of prenatal maternal stressors on fetal exposure to glucocorticoids is placental metabolism of glucocorticoids via the 11β -hydroxysteroid dehydrogenase enzyme (11β -HSD)-1 and -2. Experimental studies demonstrate that blocking activity of the 11β -HSD-2 isozyme, thereby exposing the fetus to increased concentrations of maternal glucocorticoid, has significant effects on the fetal HPA axis that persist into postnatal life [151]. Maternal undernutrition also affects placental 11β -HSD-2 activity [149,152]; it is less clear whether nutritionally induced downregulation of 11β -HSD-2 activity is accompanied by exposure of the fetus to increased glucocorticoid concentrations [153]. As a result, glucocorticoids may mediate some of the effects of maternal stressors, but other factors, such as specific nutrients or other hormonal changes, alter maternal adaptation to pregnancy [154] and, therefore, may also be involved.

Thus, maternal undernutrition may affect the materno–placental HPA axis to mediate effects on the fetus, as well as affect the fetal HPA axis directly. Maternal undernutrition affects mRNA expression levels of corticotrophin-releasing hormone in the hypothalamus, *GR* and *POMC* in the pituitary, as well as *ACTH* receptor in adrenal gland [155]. These effects may alter HPA sensitivity to acute and chronic stressors, as well as HPA axis feedback

regulatory systems [138]. As a result, a suboptimal environment *in utero* may alter both the set point and sensitivity of the fetal HPA axis [138]. Such alterations in HPA axis function are likely to play an important role in the gestational modifiers of disease risk in adult life [147,156].

Maternal undernutrition in rats [149], guinea pigs [76] and sheep [157,158] results in altered basal glucocorticoid concentrations in late fetal or postnatal life. In sheep, moderate undernutrition only in early pregnancy has been shown to result in increased fetal adrenal P450c17 protein expression in late gestation [159] and a precocious fetal cortisol surge leading to preterm birth [55]. Periconceptional and gestational undernutrition in sheep results in elevated fetal plasma ACTH concentrations, and upregulation of mRNA expression of the ACTH receptor (*MC2R*) and *StAR* in the adrenal gland of near-term ovine fetuses [160].

In addition to effects on the fetal adrenal gland, late-gestation fetal ACTH responses to metyrapone (which blocks adrenal cortisol production) demonstrate that central HPA feedback sensitivity is also affected by periconceptional undernutrition [70,158]. These effects are likely to be mediated by alterations in levels of the glucocorticoid and/or mineralocorticoid receptors in the brain, as has been demonstrated in fetal guinea pigs following maternal undernutrition [148] by altered regulation of local glucocorticoid concentrations, by effects on the 11 β -HSD isozymes [161,162] or by epigenetic changes in the hypothalamus [163,164].

Once again, fetal perturbations of HPA axis function persist into postnatal life. Maternal undernutrition in late gestation led to increased mineralocorticoid-to-glucocorticoid receptor mRNA expression ratio in the hippocampus, as well as greater *POMC* mRNA expression in the anterior pituitary of 8-month-old rat offspring [165], despite high basal plasma levels of corticosterone. Mineralocorticoid receptor binding capacities in the anterior pituitary gland were decreased, as was the mRNA expression of 11 β -HSD isozymes in the pituitary gland [166]. By contrast, *11 β -HSD-1* gene expression in the hippocampus was increased [166]. These data indicate both HPA axis hyperactivity and altered glucocorticoid negative feedback [166]. Thus, the effects of maternal undernutrition on HPA feedback sensitivity seen in fetal life can also be demonstrated in postnatal life, potentially leading to the development of adult diseases via persistent glucocorticoid excess [166].

The postnatal effects of exposure to maternal undernutrition on the HPA axis also change with postnatal age. Undernutrition of ewes in the first month of gestation led to a significant increase in resting plasma cortisol of female offspring at 12 months [77]. Similarly, early gestational undernutrition led to greater ACTH and cortisol response to corticotropin-releasing hormone challenge in the offspring at 2 months and higher basal cortisol levels at 5.5 months [167]. However, periconceptional undernutrition results in suppressed HPA axis responses to corticotrophic stimulation and to isolation stress in male offspring, which worsens with advancing age [168]. A similar suppression of HPA axis function with increasing age is seen in sheep exposed to exogenous glucocorticoids *in utero* [169,170]. This pattern of

advanced maturation in the fetus, followed by suppressed or impaired function in adult postnatal life, is consistent with that seen in pancreatic function.

Therefore, the HPA axis may play a critical role in the postnatal endocrine effects of maternal undernutrition. The effects on fetal endocrine development may be mediated, at least in part, by exposure of the developing fetal organs to altered circulating or tissue concentrations of glucocorticoids. Furthermore, altered postnatal HPA function may mediate many of the adverse health effects seen in the offspring of mothers undernourished during pregnancy.

Somatotrophic axis

The fetal somatotrophic axis is functional *in utero* [171] and, during prenatal development, GH acts on many tissues at all developmental stages, although it appears to have a relatively minor influence on fetal growth [172,173]. IGF-1 and IGF-2 are the hormones primarily regulating fetal growth, acting in both paracrine and endocrine fashions [174–176]. IGF-2 is thought to be most important in embryonic and early gestational growth [177], while IGF-1 is considered the main endocrine growth regulator in late gestation [178,179].

The fetal somatotrophic axis is substantially regulated by the nutritional status of the mother [171]. Maternal undernutrition reduces fetal plasma concentrations of IGF-1 and IGF-2, and reduces their mRNA expression in a number of tissues [180]. In sheep, this reduction in fetal plasma IGF-1 concentrations appears to occur throughout gestation, and has been demonstrated after maternal undernutrition from early-to-mid gestation [157], early-to-late gestation [181] and mid-to-late gestation [182]. More detailed studies in chronically catheterized sheep have shown that fetal plasma IGF-1 concentrations are regulated by fetal glucose and insulin but not amino acid concentrations [183,184], and that this is the probable mechanism by which maternal nutritional status regulates fetal growth [185].

Fetal plasma concentrations of IGF binding proteins (IGFBPs) also have been shown to be nutritionally regulated [186]. In rats, maternal undernutrition increased fetal plasma IGFBP-1 and IGFBP-2 concentrations, thus restricting the availability of circulating IGF-1. Similarly, IGFBP-2 concentrations were higher in the fetuses of undernourished ewes [181], as were *IGF-1R* and *IGF-2R* mRNA expression in fetal adipose tissue at term [49].

There is also some evidence that these nutritionally induced changes in the IGF axis are prolonged beyond the period of maternal undernutrition. Fetuses of ewes undernourished only in the periconceptional period showed enhanced responsiveness of IGF-1, IGFBP-1 and IGFBP-3 to an undernutrition insult in late gestation [187]. In rats, plasma IGF-1 was reduced and IGFBP-1 and -2 increased in early postnatal life in the offspring of undernourished dams [188].

Growth hormone is also nutritionally regulated both before and after birth. Fetuses of ewes severely undernourished in late gestation had higher GH concentrations and GH mass secreted per burst, as well as increased GH peak and nadir levels [171]. In addition, in sheep, the number of GH secretory episodes increased,

while the response to an exogenous GnRH challenge was significantly lower in male nutrient-restricted fetuses [189]. After birth, sheep that had experienced fetal undernutrition as a result of placental restriction had increased plasma GH concentrations in pubertal and adult males, but lower levels in adult females [190].

Human data on the postnatal effects of prenatal undernutrition are conflicting. SGA children have been shown to have lower serum IGF-1 levels and higher overnight GH secretion, with an apparent shift in the set point of their GH–IGF-1 axis [191], consistent with other reports that IUGR children are partially IGF-1 resistant [192]. However, prepubertal children with short stature after IUGR have also been reported to have decreased GH secretion due to impaired GH pulse amplitudes, as well as having reduced plasma IGF-1 and IGF-2 concentrations, probably indicating GH insufficiency [193]. Boys (but not girls) also had reduced overnight urinary GH excretion [193]. Similarly, both low birthweight and low placental weight were associated with reduced GH excretion in men and women at 20 years of age [194].

Such changes may be the result of permanent effects in the hypothalamus, as adult growth-restricted-born rats showed altered mRNA expression in the hypothalamus, with increased somatostatin in the periventricular nucleus and decreased NPY in the arcuate nucleus [195,196]. However, these alterations were sex specific, with male but not female rats showing decreased galanin mRNA expression in GHRH neurons that control GH secretion [197].

Hypothalamic–pituitary–thyroid axis

Thyroid hormones play important roles in embryogenesis and fetal maturation [198], having a crucial role in the development of the fetal CNS in all mammals, including humans [199,200]. Maternal undernutrition in late gestation in rats led to decreased fetal plasma T_4 concentrations [148], as did maternal undernutrition throughout gestation in sheep [189]. However, others have reported little effect of maternal undernutrition on fetal plasma thyroid concentrations in sheep [182] and guinea pigs [201]. After birth, lambs born with IUGR following placental restriction had persistently reduced total T_4 and elevated total T_3 plasma concentrations [202]. These changes may reflect altered tissue hormone conversion, since elevated mRNA expression of the prohormone deiodinase type 2 was found in the hippocampus and cerebellum of fetuses of undernourished guinea pigs [203]. Again, the effects appear to be sex-specific, with increased *TR- α* and *- β* mRNA expression in male brains, but decreased expression levels in female brains [203].

Plasma thyroid hormone concentrations are also reduced in IUGR human fetuses, while thyroid-stimulating hormone (TSH) levels are increased [204], as are *TR- α* and *TR- β* mRNA expression in the placenta [205]. However, there is reduced expression of all thyroid receptor isoform proteins in the cerebral cortex and cerebellum of severe IUGR human fetuses [206]. By contrast, another study showed that, at birth, cord blood T_4 concentrations were higher, while recombinant T_3 concentrations were lower in neonates born to malnourished and/or

anemic mothers [28,207]. Nonetheless, postnatal serum T_3 and T_4 concentrations were reduced in SGA babies in the first few weeks after birth [208,209].

Maternal undernutrition in rats during late gestation and lactation led to considerable reductions in plasma concentrations of free T_3 , free T_4 and TSH in 14-day-old offspring, as well as histological changes and weight reduction in the thyroid gland [210]. Maternal refeeding early in postnatal life (4 days) restored thyroid weight and plasma thyroid hormone concentrations to control levels at 14 days, but plasma TSH failed to fully recover [210]. Similarly, protein restriction during lactation in rats led to hypothyroidism, and although refeeding restored serum T_3 , T_4 and TSH to normal levels by 60 days, there was continuing high radioiodine uptake, suggesting permanent changes in thyroid function [211]. Indeed, a subsequent study showed that the same undernutrition regime led to hyperthyroidism in the adult offspring [212]. Thus, in rats, lactation seems to be a critical period for long-term effects of maternal undernutrition on thyroid function [213].

However, there is a paucity of data on the long-term effects of maternal undernutrition on the hypothalamic–pituitary–thyroid axis, and most of the available data refer to the effects of deficiency of iodine, which is an essential component of T_3 and T_4 . The fetal supply of T_4 during the first half of human pregnancy is primarily of maternal origin, so that maternal iodine deficiency may have severe and irreversible effects on fetal neurological development [198,214] and permanent neurological impairment in the neonate offspring [198,214,215].

Conclusion

Maternal undernutrition, whether it occurs before conception, during gestation or during lactation, may alter endocrine development in the offspring, ultimately affecting health in later life. However, the effects of the maternal undernutrition insult vary according to its timing, severity, duration and nature. Fetal number and gender also influence fetal endocrine development and the long-term effects of maternal undernutrition. Importantly, effects of maternal undernutrition may evolve over the lifetime of the individual and, therefore, are carried over into subsequent generations.

Clearly, the effects of maternal undernutrition on the endocrine development of the offspring are complex, and involve multiple inter-related mechanisms. Epigenetic regulation of gene transcription is likely to be one important mechanism, but many aspects are still obscure. Since poor maternal and child nutrition is common in both developing and developed countries [216,217], it is paramount that we not only better understand the mechanisms underlying its long-term effects, but also identify treatments to ameliorate the associated mortality and morbidity.

Expert commentary

In view of the numerous effects on the offspring endocrine development associated with maternal undernutrition, it is desirable to develop appropriate preventive strategies to minimize both incidence and severity of associated long-term health effects. Improving the nutritional status of women prior to and during

pregnancy would be an obvious intervention. However, such measures would have to be appropriately targeted to specific groups, based on cultural, social and economic status, which constitutes a considerable public health challenge.

Other possible clinical strategies include nutrient supplementation, and at least one study has shown that folic acid supplementation during pregnancy prevented epigenetic modification of hepatic gene expression in the offspring of protein-restricted rats [218]. Furthermore, additional research is needed into postnatal interventions that can reverse the alterations in the endocrine system induced by maternal undernutrition. Leptin treatment in early postnatal life, for example, normalized the undernutrition-induced phenotype in adult rats, indicating that some prenatal adaptations resulting from fetal undernutrition can be reversed postnatally [113].

Five-year view

In the next 5 years, we will understand the epigenetic mechanisms, use them as diagnostic markers and be looking at early postnatal interventions based on reversing the nutritional and endocrine consequences of prenatal events. Public health focus will turn to optimal nutrition of women before pregnancy and in the offspring in early postnatal life.

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Key issues

- Maternal undernutrition affects the endocrine development of the offspring, not only when it occurs during gestation but also during the preconceptional period and during lactation.
- Both micro- and macro-nutrient restriction may have long-term effects.
- The effects vary according to its timing, severity, duration and nature of the undernutrition.
- The effects also vary according to the sex of the offspring.
- In some cases, accelerated fetal maturation of the endocrine axis is associated with early activation but later suppression of that axis in postnatal life.
- The hypothalamic–pituitary–adrenal axis is one of several endocrine axes involved but the effects cannot be explained by simple exposure to excess glucocorticoids.
- Epigenetic mechanisms may underlie some of the long-term effects of maternal undernutrition.

References

- Barker DJP. *Mothers, Babies and Disease in Later Life*. BMJ Publishing Group, London, UK (1994).
- Wintour EM, Owens JA. *Early Life Origins of Health and Disease*. Springer, NY, USA (2006).
- Bateson P, Barker D, Clutton-Brock T *et al*. Developmental plasticity and human health. *Nature* 430 (6998), 419–421 (2004).
- Weinhold B. Epigenetics: the science of change. *Environ. Health Perspect.* 114(3), 160–167 (2006).
- Wu G, Bazer FW, Cudd TA, Meininger CJ, Spencer TE. Maternal nutrition and fetal development. *J. Nutr.* 134(9), 2169–2172 (2004).
- Lillicrop KA, Hanson MA, Burdge GC. Epigenetics and the influence of maternal diet. In: *Early Life Origins of Human Health and Disease*. Newnham JP, Ross MG (Eds). Karger, Basel, Switzerland 11–20 (2009).
- Kim Y-I. Nutritional epigenetics: impact of folate deficiency on DNA methylation and colon cancer susceptibility. *J. Nutr.* 135(11), 2703–2709 (2005).
- Waterland RA, Jirtle RL. Transposable elements: targets for early nutritional effects on epigenetic gene regulation. *Mol. Cell. Biol.* 23(15), 5293–5300 (2003).
- Egger G, Liang G, Aparicio A, Jones PA. Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 429(6990), 457–463 (2004).
- Epigenetic Markers*. Widschwendter M (Ed.). IOS Press, Amsterdam, The Netherlands (2007).
- Gluckman PD, Hanson MA, Cooper C, Thornburg KL. Effect of *in utero* and early-life conditions on adult health and disease. *N. Engl. J. Med.* 359(1), 61–73 (2008).
- Jones HN, Woollett LA, Barbour N, Prasad PD, Powell TL, Jansson T. High-fat diet before and during pregnancy causes marked up-regulation of placental nutrient transport and fetal overgrowth in C57/BL6 mice. *FASEB J.* 23(1), 271–278 (2009).
- Armitage JA, Taylor PD, Poston L. Experimental models of developmental programming: consequences of exposure to an energy rich diet during development. *J. Physiol.* 565(1), 3–8 (2005).
- Lawlor DA, Timpson NJ, Harbord RM *et al*. Exploring the developmental overnutrition hypothesis using parental-offspring associations and *FTO* as an instrumental variable. *PLoS Med.* 5(3), E33 (2008).
- Waller DK, Dawson TE. Relationship between maternal obesity and adverse pregnancy outcomes. In: *The Impact of Maternal Nutrition on the Offspring*. Hornstra G, Uauy R, Yang X (Eds). Karger, Basel, Switzerland 197–212 (2005).
- King JC. Maternal obesity, metabolism, and pregnancy outcomes. *Annu. Rev. Nutr.* 26, 271–291 (2006).
- Chen H, Simar D, Lambert K, Mercier J, Morris MJ. Maternal and postnatal overnutrition differentially impact appetite regulators and fuel metabolism. *Endocrinology* 149(11), 5348–5356 (2008).
- Taylor PD, Poston L. Developmental programming of obesity in mammals. *Exp. Physiol.* 92(2), 287–298 (2007).
- Levin BE. Metabolic imprinting: critical impact of the perinatal environment on the regulation of energy homeostasis. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 361(1471), 1107–1121 (2006).

- 20 Catalano PM, Ehrenberg HM. The short- and long-term implications of maternal obesity on the mother and her offspring. *BJOG* 113(10), 1126–1133 (2006).
- 21 Black RE. Micronutrients in pregnancy. *Br. J. Nutr.* 85(Suppl. 2), S193–S197 (2001).
- 22 Godfrey KM, Barker DJ. Fetal nutrition and adult disease. *Am. J. Clin. Nutr.* 71(5 Suppl.), 1344–1352 (2000).
- 23 Roseboom T, de Rooij S, Painter R. The Dutch famine and its long-term consequences for adult health. *Early Hum. Dev.* 82(8), 485–491 (2006).
- 24 Godfrey K, Robinson S, Barker DJ, Osmond C, Cox V. Maternal nutrition in early and late pregnancy in relation to placental and fetal growth. *BMJ* 312(7028), 410–414 (1996).
- 25 Rao S, Yajnik CS, Kanade A *et al.* Intake of micronutrient-rich foods in rural Indian mothers is associated with the size of their babies at birth: Pune Maternal Nutrition Study. *J. Nutr.* 131(4), 1217–1224 (2001).
- 26 Venu L, Harishankar N, Prasanna KT, Raghunath M. Maternal dietary vitamin restriction increases body fat content but not insulin resistance in WNIN rat offspring up to 6 months of age. *Diabetologia* 47(9), 1493–1501 (2004).
- 27 Gambling L, Dunford S, Wallace DI *et al.* Iron deficiency during pregnancy affects postnatal blood pressure in the rat. *J. Physiol.* 552(2), 603–610 (2003).
- 28 Mahajan S, Aalinkel R, Shah P, Singh S, Kochupillai N. Nutritional anaemia dysregulates endocrine control of fetal growth. *Br. J. Nutr.* 100(2), 408–417 (2008).
- 29 Dunn JT. Iodine supplementation and the prevention of cretinism. *Ann. NY Acad. Sci.* 678, 158–168 (1993).
- 30 Tomat AL, Inserra F, Veiras L *et al.* Moderate zinc restriction during fetal and postnatal growth of rats: effects on adult arterial blood pressure and kidney. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 295(2), R543–R549 (2008).
- 31 Bergel E, Belizán JM. A deficient maternal calcium intake during pregnancy increases blood pressure of the offspring in adult rats. *BJOG* 109(5), 540–545 (2002).
- 32 Belizan JM, Villar J, Bergel E *et al.* Long-term effect of calcium supplementation during pregnancy on the blood pressure of offspring: follow up of a randomised controlled trial. *BMJ* 315(7103), 281–285 (1997).
- 33 Hiller JE, Crowther CA, Moore VA, Willson K, Robinson JS. Calcium supplementation in pregnancy and its impact on blood pressure in children and women: follow up of a randomised controlled trial. *Aust. NZ J. Obstet. Gynaecol.* 47(2), 115–121 (2007).
- 34 Langley SC, Jackson AA. Increased systolic blood pressure in adult rats induced by fetal exposure to maternal low protein diets. *Clin. Sci.* 86(2), 217–222 (1994).
- 35 Skilton MR, Gosby AK, Wu BJ *et al.* Maternal undernutrition reduces aortic wall thickness and elastin content in offspring rats without altering endothelial function. *Clin. Sci.* 111(4), 281–287 (2006).
- 36 Cheema KK, Dent MR, Saini HK, Aroutiounova N, Tappia PS. Prenatal exposure to maternal undernutrition induces adult cardiac dysfunction. *Br. J. Nutr.* 93(4), 471–477 (2005).
- 37 Gangula PRR, Reed L, Yallampalli C. Antihypertensive effects of flutamide in rats that are exposed to a low-protein diet *in utero*. *Am. J. Obstet. Gynecol.* 192(3), 952–960 (2005).
- 38 Adair LS, Kuzawa CW, Borja J. Maternal energy stores and diet composition during pregnancy program adolescent blood pressure. *Circulation* 104(9), 1034–1039 (2001).
- 39 Huh SY, Rifas-Shiman SL, Kleinman KP, Rich-Edwards JW, Lipshultz SE, Gillman MW. Maternal protein intake is not associated with infant blood pressure. *Int. J. Epidemiol.* 34(2), 378–384 (2005).
- 40 Sathishkumar K, Elkins R, Yallampalli U, Yallampalli C. Protein restriction during pregnancy induces hypertension and impairs endothelium-dependent vascular function in adult female offspring. *J. Vasc. Res.* 46, 229–239 (2009).
- 41 Bellinger L, Sculley DV, Langley-Evans SC. Exposure to undernutrition in fetal life determines fat distribution, locomotor activity and food intake in ageing rats. *Int. J. Obes.* 30(5), 729–738 (2006).
- 42 Langley SC, Browne RF, Jackson AA. Altered glucose tolerance in rats exposed to maternal low protein diets *in utero*. *Comp. Biochem. Physiol. A Physiol.* 109, 223–229 (1994).
- 43 Zambrano E, Bautista CJ, Deás M *et al.* A low maternal protein diet during pregnancy and lactation has sex- and window of exposure-specific effects on offspring growth and food intake, glucose metabolism and serum leptin in the rat. *J. Physiol.* 571(1), 221–230 (2006).
- 44 Yates Z, Tarling EJ, Langley-Evans SC, Salter AM. Maternal undernutrition programmes atherosclerosis in the *ApoE*3-Leiden* mouse. *Br. J. Nutr.* 101(8), 1185–1194 (2009).
- 45 Petry CJ, Ozanne SE, Wang CL, Hales CN. Early protein restriction and obesity independently induce hypertension in 1-year-old rats. *Clin. Sci.* 93(2), 147–152 (1997).
- 46 Fernandez-Twinn DS, Wayman A, Ekizoglou S, Martin MS, Hales CN, Ozanne SE. Maternal protein restriction leads to hyperinsulinemia and reduced insulin-signaling protein expression in 21-mo-old female rat offspring. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 288(2), R368–R373 (2005).
- 47 Ozanne SE, Nicholas Hales C. Poor fetal growth followed by rapid postnatal catch-up growth leads to premature death. *Mech. Ageing Dev.* 126(8), 852–854 (2005).
- 48 Roseboom TJ, van der Meulen JH, Osmond C, Barker DJ, Ravelli AC, Bleker OP. Plasma lipid profiles in adults after prenatal exposure to the Dutch famine. *Am. J. Clin. Nutr.* 72(5), 1101–1106 (2000).
- 49 Symonds ME, Pearce S, Bispham J, Gardner DS, Stephenson T. Timing of nutrient restriction and programming of fetal adipose tissue development. *Proc. Nutr. Soc.* 63(3), 397–403 (2004).
- 50 Joshi S, Garole V, Daware M, Girigosavi S, Rao S. Maternal protein restriction before pregnancy affects vital organs of offspring in Wistar rats. *Metabolism* 52(1), 13–18 (2003).
- 51 Kwong WY, Wild AE, Roberts P, Willis AC, Fleming TP. Maternal undernutrition during the preimplantation period of rat development causes blastocyst abnormalities and programming of postnatal hypertension. *Development* 127, 4195–4202 (2000).
- 52 Watkins AJ, Ursell E, Pantou R *et al.* Adaptive responses by mouse early embryos to maternal diet protect fetal growth but predispose to adult onset disease. *Biol. Reprod.* 78(2), 299–306 (2008).
- 53 Oliver MH, Hawkins P, Harding JE. Periconceptional undernutrition alters growth trajectory, endocrine and metabolic responses to fasting in late gestation fetal sheep. *Pediatr. Res.* 57(4), 591–598 (2005).
- 54 Rumball CW, Bloomfield FH, Oliver MH, Harding JE. Different periods of periconceptional undernutrition have different effects on growth, metabolic and endocrine status in fetal sheep. *Pediatr. Res.* 66(6), 605–613 (2009).

- 55 Bloomfield FH, Oliver MH, Hawkins P *et al.* A periconceptual nutritional origin for noninfectious preterm birth. *Science* 300(5619), 606 (2003).
- 56 Oliver MH, Hawkins P, Breier BH, Van Zijl PL, Sargison SA, Harding JE. Maternal undernutrition during the periconceptual period increases plasma taurine levels and insulin response to glucose but not arginine in the late gestational fetal sheep. *Endocrinology* 142(10), 4576–4579 (2001).
- 57 Todd SE, Oliver MH, Jaquiere AL, Bloomfield FH, Harding JE. Periconceptual undernutrition of ewes impairs glucose tolerance in their adult offspring. *Pediatr. Res.* 65(4), 409–413 (2009).
- 58 Hernandez CE, Harding JE, Oliver MH, Bloomfield FH, Held SDE, Matthews LR. Effects of litter size, sex and periconceptual ewe nutrition on side preference and cognitive flexibility in the offspring. *Behav. Brain Res.* 120(1–2), 76–83 (2009).
- 59 Lea RG, Andrade LP, Rae MT *et al.* Effects of maternal undernutrition during early pregnancy on apoptosis regulators in the ovine fetal ovary. *Reproduction* 131(1), 113–124 (2006).
- 60 Kotsampasi B, Chadio S, Papadomichelakis G *et al.* Effects of maternal undernutrition on the hypothalamic–pituitary–gonadal axis function in female sheep offspring. *Reprod. Domest. Anim.* 44(4), 677–684 (2009).
- 61 Budge H, Gnanalingham MG, Gardner DS, Mostyn A, Stephenson T, Symonds ME. Maternal nutritional programming of fetal adipose tissue development: long-term consequences for later obesity. *Birth Defects Res. C. Embryo Today* 75(3), 193–199 (2005).
- 62 Ravelli AC, van der Meulen JH, Michels RP *et al.* Glucose tolerance in adults after prenatal exposure to famine. *Lancet* 351(9097), 173–177 (1998).
- 63 Gardner DS, Tingey K, Van Bon BWM *et al.* Programming of glucose–insulin metabolism in adult sheep after maternal undernutrition. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 289(4), R947–R954 (2005).
- 64 Bloomfield FH, Oliver MH, Giannoulas CD, Gluckman PD, Harding JE, Challis JR. Brief undernutrition in late-gestation sheep programs the hypothalamic–pituitary–adrenal axis in adult offspring. *Endocrinology* 144(7), 2933–2940 (2003).
- 65 Budge H, Dandrea J, Mostyn A *et al.* Differential effects of fetal number and maternal nutrition in late gestation on prolactin receptor abundance and adipose tissue development in the neonatal lamb. *Pediatr. Res.* 53(2), 302–308 (2003).
- 66 Brennan KA, Gopalakrishnan GS, Kurlak L *et al.* Impact of maternal undernutrition and fetal number on glucocorticoid, growth hormone and insulin-like growth factor receptor mRNA abundance in the ovine fetal kidney. *Reproduction* 129(2), 151–159 (2005).
- 67 Edwards LJ, McMillen IC. Impact of maternal undernutrition during the periconceptual period, fetal number, and fetal sex on the development of the hypothalamo–pituitary adrenal axis in sheep during late gestation. *Biol. Reprod.* 66, 1562–1569 (2002).
- 68 MacLaughlin SM, Walker SK, Kleemann DO *et al.* Impact of periconceptual undernutrition on adrenal growth and adrenal insulin-like growth factor and steroidogenic enzyme expression in the sheep fetus during early pregnancy. *Endocrinology* 148(4), 1911–1920 (2007).
- 69 Rumball CW, Harding JE, Oliver MH, Bloomfield FH. Effects of twin pregnancy and periconceptual undernutrition on maternal metabolism, fetal growth and glucose–insulin axis function in ovine pregnancy. *J. Physiol.* 586(5), 1399–1411 (2008).
- 70 Rumball CW, Oliver MH, Thorstensen EB *et al.* Effects of twinning and periconceptual undernutrition on late-gestation hypothalamic–pituitary–adrenal axis function in ovine pregnancy. *Endocrinology* 149(3), 1163–1172 (2008).
- 71 Jones AP, Assimon SA, Friedman MI. The effect of diet on food intake and adiposity in rats made obese by gestational undernutrition. *Physiol. Behav.* 37(3), 381–386 (1986).
- 72 Jones AP, Simson EL, Friedman MI. Gestational undernutrition and the development of obesity in rats. *J. Nutr.* 114(8), 1484–1492 (1984).
- 73 Anguita RM, Sigulem DM, Sawaya AL. Intrauterine food restriction is associated with obesity in young rats. *J. Nutr.* 123, 1421–1428 (1993).
- 74 McMullen S, Langley-Evans SC. Sex-specific effects of prenatal low-protein and carbenoxolone exposure on renal angiotensin receptor expression in rats. *Hypertension* 46(6), 1374–1380 (2005).
- 75 Hoppe CC, Evans RG, Bertram JF, Moritz KM. Effects of dietary protein restriction on nephron number in the mouse. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 292(5), R1768–R1774 (2007).
- 76 Lingas RI, Matthews SG. A short period of maternal nutrient restriction in late gestation modifies pituitary–adrenal function in adult guinea pig offspring. *Neuroendocrinology* 73(5), 302–311 (2001).
- 77 Gardner DS, Van Bon BWM, Dandrea J *et al.* Effect of periconceptual undernutrition and gender on hypothalamic–pituitary–adrenal axis function in young adult sheep. *J. Endocrinol.* 190(2), 203–212 (2006).
- 78 Ojeda NB, Grigore D, Robertson EB, Alexander BT. Estrogen protects against increased blood pressure in postpubertal female growth restricted offspring. *Hypertension* 50(4), 679–685 (2007).
- 79 Foecking EM, McDevitt MA, Acosta-Martínez M, Horton TH, Levine JE. Neuroendocrine consequences of androgen excess in female rodents. *Horm. Behav.* 53(5), 673–692 (2008).
- 80 Abdel-Hakeem AK, Henry TQ, Magee TR *et al.* Mechanisms of impaired nephrogenesis with fetal growth restriction: altered renal transcription and growth factor expression. *Am. J. Obstet. Gynecol.* 199(3), E251–E252 (2008).
- 81 Brennan KA, Kaufman S, Reynolds SW *et al.* Differential effects of maternal nutrient restriction through pregnancy on kidney development and later blood pressure control in the resulting offspring. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 295(1), R197–R205 (2008).
- 82 Welham SJJ, Riley PR, Wade A, Hubank M, Woolf AS. Maternal diet programs embryonic kidney gene expression. *Physiol. Genomics* 22(1), 48–56 (2005).
- 83 Henry T, Torday J, Magee T *et al.* Maternal food restriction inhibits nephrogenesis by disrupting mesonephric mesenchyme ureteric bud signaling [conference abstract]. *Early Hum. Dev.* 83(Suppl. 1), S176 (2007).
- 84 Woods LL, Weeks DA, Rasch R. Programming of adult blood pressure by maternal protein restriction: role of nephrogenesis. *Kidney Int.* 65(4), 1339–1348 (2004).
- 85 Gilbert JS, Lang AL, Grant AR, Nijland MJ. Maternal nutrient restriction in sheep: hypertension and decreased nephron number in offspring at 9 months of age. *J. Physiol.* 565(1), 137–147 (2005).

- 86 Painter RC, Roseboom TJ, van Montfrans GA *et al.* Microalbuminuria in adults after prenatal exposure to the dutch famine. *J. Am. Soc. Nephrol.* 16, 189–194 (2005).
- 87 Chen C-M, Chou H-S. Effects of maternal undernutrition on glomerular ultrastructure in rat offspring. *Pediatr. Neonatol.* 50(2), 50–53 (2009).
- 88 Khorram O, Khorram N, Momeni M *et al.* Maternal undernutrition inhibits angiogenesis in the offspring: a potential mechanism of programmed hypertension. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 293(2), R745–R753 (2007).
- 89 Keller G, Zimmer G, Mall G, Ritz E, Amann K. Nephron number in patients with primary hypertension. *N. Engl. J. Med.* 348(2), 101–108 (2003).
- 90 Woods LL, Ingelfinger JR, Nyengaard JR, Rasch R. Maternal protein restriction suppresses the newborn renin-angiotensin system and programs adult hypertension in rats. *Pediatr. Res.* 49(4), 460–467 (2001).
- 91 Vehaskari VM, Stewart T, Lafont D, Soyze C, Seth D, Manning J. Kidney angiotensin and angiotensin receptor expression in prenatally programmed hypertension. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 287(2), F262–F267 (2004).
- 92 Sahajpal V, Ashton N. Increased glomerular angiotensin II binding in rats exposed to a maternal low protein diet *in utero*. *J. Physiol.* 563(1), 193–201 (2005).
- 93 Sahajpal V, Ashton N. Renal function and angiotensin AT₁ receptor expression in young rats following intrauterine exposure to a maternal low-protein diet. *Clin. Sci.* 104(6), 607–614 (2003).
- 94 Sherman RC, Langley-Evans SC. Antihypertensive treatment in early postnatal life modulates prenatal dietary influences upon blood pressure in the rat. *Clin. Sci.* 98(3), 269–275 (2000).
- 95 Ravelli AC, van Der Meulen JH, Osmond C, Barker DJ, Bleker OP. Obesity at the age of 50 y in men and women exposed to famine prenatally. *Am. J. Clin. Nutr.* 70(5), 811–816 (1999).
- 96 Ford SP, Hess BW, Schwowe MM *et al.* Maternal undernutrition during early to mid-gestation in the ewe results in altered growth, adiposity, and glucose tolerance in male offspring. *J. Anim. Sci.* 85(5), 1285–1294 (2007).
- 97 Bispham J, Gopalakrishnan GS, Dandrea J *et al.* Maternal endocrine adaptation throughout pregnancy to nutritional manipulation: consequences for maternal plasma leptin and cortisol and the programming of fetal adipose tissue development. *Endocrinology* 144(8), 3575–3585 (2003).
- 98 Bispham J, Gardner DS, Gnanalingham MG, Stephenson T, Symonds ME, Budge H. Maternal nutritional programming of fetal adipose tissue development: differential effects on messenger ribonucleic acid abundance for uncoupling proteins and peroxisome proliferator-activated and prolactin receptors. *Endocrinology* 146(9), 3943–3949 (2005).
- 99 Mostyn A, Wilson V, Dandrea J *et al.* Ontogeny and nutritional manipulation of mitochondrial protein abundance in adipose tissue and the lungs of postnatal sheep. *Br. J. Nutr.* 90(2), 323–328 (2003).
- 100 Voehringer DW, Hirschberg DL, Xiao J *et al.* Gene microarray identification of redox and mitochondrial elements that control resistance or sensitivity to apoptosis. *Proc. Natl Acad. Sci. USA* 97(6), 2680–2685 (2000).
- 101 Symonds ME, Lomax MA. Maternal and environmental influences on thermoregulation in the neonate. *Proc. Nutr. Soc.* 51(2), 165–172 (1992).
- 102 Symonds ME, Stephenson T. Maternal nutrient restriction and endocrine programming of fetal adipose tissue development. *Biochem. Soc. Trans.* 27, 97–103 (1999).
- 103 Lorenzo M, Valverde AM, Teruel T, Benito M. IGF-I is a mitogen involved in differentiation-related gene expression in fetal rat brown adipocytes. *J. Cell Biol.* 123(6), 1567–1575 (1993).
- 104 Yuen BS, McMillen IC, Symonds ME, Owens PC. Abundance of leptin mRNA in fetal adipose tissue is related to fetal body weight. *J. Endocrinol.* 163(3), R11–R14 (1999).
- 105 Smith JT, Waddell BJ. Leptin distribution and metabolism in the pregnant rat: transplacental leptin passage increases in late gestation but is reduced by excess glucocorticoids. *Endocrinology* 144(7), 3024–3030 (2003).
- 106 Yura S, Itoh H, Sagawa N *et al.* Neonatal exposure to leptin augments diet-induced obesity in leptin-deficient Ob/Ob mice. *Obesity* 16(6), 1289–1295 (2008).
- 107 Grattan DR. Fetal programming from maternal obesity: eating too much for two? *Endocrinology*, 149(11), 5345–5347 (2008).
- 108 Gopalakrishnan GS, Gardner DS, Rhind SM *et al.* Programming of adult cardiovascular function after early maternal undernutrition in sheep. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 287(1), R12–R20 (2004).
- 109 Sebert SP, Hyatt MA, Chan LLY *et al.* Maternal nutrient restriction between early and midgestation and its impact upon appetite regulation after juvenile obesity. *Endocrinology* 150(2), 634–641 (2009).
- 110 Yura S, Itoh H, Sagawa N *et al.* Role of premature leptin surge in obesity resulting from intrauterine undernutrition. *Cell Metab.* 1(6), 371–378 (2005).
- 111 Breton C, Lukaszewski M-A, Risold P-Y *et al.* Maternal prenatal undernutrition alters the response of POMC neurons to energy status variation in adult male rat offspring. *Am. J. Physiol. Endocrinol. Metab.* 296(3), E462–E472 (2009).
- 112 Delahaye F, Breton C, Risold P-Y *et al.* Maternal perinatal undernutrition drastically reduces postnatal leptin surge and affects the development of arcuate nucleus proopiomelanocortin neurons in neonatal male rat pups. *Endocrinology* 149(2), 470–475 (2008).
- 113 Vickers MH, Gluckman PD, Coveny AH *et al.* Neonatal leptin treatment reverses developmental programming. *Endocrinology* 146(10), 4211–4216 (2005).
- 114 Rhind SM. Effects of maternal nutrition on fetal and neonatal reproductive development and function. *Anim. Reprod. Sci.* 82–83, 169–181 (2004).
- 115 Lumey LH, Stein AD. Offspring birth weights after maternal intrauterine undernutrition: a comparison within sibships. *Am. J. Epidemiol.* 146(10), 810–819 (1997).
- 116 Zambrano E, Martínez-Samayoa PM, Bautista CJ *et al.* Sex differences in transgenerational alterations of growth and metabolism in progeny (F₂) of female offspring (F₁) of rats fed a low protein diet during pregnancy and lactation. *J. Physiol.* 566(1), 225–236 (2005).
- 117 Zambrano E. [The transgenerational mechanisms in developmental programming of metabolic diseases]. *Rev. Invest. Clin.* 61(1), 41–52 (2009).
- 118 Rae MT, Rhind SM, Fowler PA, Miller DW, Kyle CE, Brooks AN. Effect of maternal undernutrition on fetal testicular steroidogenesis during the CNS androgen-responsive period in male sheep fetuses. *Reproduction* 124(1), 33–39 (2002).
- 119 Teixeira CV, Silandre D, de Souza Santos AM *et al.* Effects of maternal undernutrition during lactation on

- aromatase, estrogen, and androgen receptors expression in rat testis at weaning. *J. Endocrinol.* 192(2), 301–311 (2007).
- 120 Sloboda DM, Hart R, Doherty DA, Pennell CE, Hickey M. Age at menarche: influences of prenatal and postnatal growth. *J. Clin. Endocrinol. Metab.* 92(1), 46–50 (2007).
- 121 Painter RC, Westendorp RGJ, de Rooij SR, Osmond C, Barker DJP, Roseboom TJ. Increased reproductive success of women after prenatal undernutrition. *Hum. Reprod.* 23(11), 2591–2595 (2008).
- 122 Da Silva P, Aitken RP, Rhind SM, Racey PA, Wallace JM. Influence of placentally mediated fetal growth restriction on the onset of puberty in male and female lambs. *Reproduction* 122(3), 375–383 (2001).
- 123 Engelbregt MJT, Houdijk ME, Popp-Snijders AC, Delemarre-Van De Waal HA. The effects of intra-uterine growth retardation and postnatal undernutrition on onset of puberty in male and female rats. *Pediatr. Res.* 48(6), 803–807 (2000).
- 124 Guzmán C, Cabrera R, Cárdenas M, Larrea F, Nathanielsz PW, Zambrano E. Protein restriction during fetal and neonatal development in the rat alters reproductive function and accelerates reproductive ageing in female progeny. *J. Physiol.* 572(1), 97–108 (2006).
- 125 Sloboda DM, Howie G, Pleasants AB, Gluckman PD, Vickers MH. Pre- and postnatal nutritional histories influence reproductive maturation and ovarian function in the rat. *PLoS ONE* 4(8), E6744 (2009).
- 126 Menendez-Patterson A, Menendez E, Fernandez S, Fernandez M, Marin B. Influence of undernutrition during gestation and suckling on development and sexual maturity in the rat. *J. Nutr.* 115(8), 1025–1032 (1985).
- 127 de Rooij SR, Painter RC, Roseboom TJ *et al.* Glucose tolerance at age 58 and the decline of glucose tolerance in comparison with age 50 in people prenatally exposed to the Dutch famine. *Diabetologia* 49(4), 637–643 (2006).
- 128 Economides DL, Nicolaides KH, Gahl WA, Bernardini I, Bottoms S, Evans M. Cordocentesis in the diagnosis of intrauterine starvation. *Am. J. Obstet. Gynecol.* 161(4), 1004 (1989).
- 129 Economides DL, Proudler A, Nicolaides KH. Plasma insulin in appropriate- and small-for-gestational-age fetuses. *Am. J. Obstet. Gynecol.* 160, 1091–1094 (1989).
- 130 Economides DL, Nicolaides KH, Gahl W, Bernardini I, Evans MI. Plasma amino acids in appropriate and small-for-gestational age infants. *Am. J. Obstet. Gynecol.* 161, 1219–1227 (1989).
- 131 Hofman PL, Cutfield WS, Robinson EM, Bergman RN, Menon RK, Gluckman PD. Insulin resistance in short children with intrauterine growth retardation. *J. Clin. Endocrinol. Metab.* 82(2), 402–406 (1997).
- 132 Leger J, Levy-Marchal C, Bloch J *et al.* Reduced final height and indications for insulin resistance in 20 year olds born small for gestational age: regional cohort study. *BMJ* 315(7104), 341–347 (1997).
- 133 Jaquet D, Gaboriau A, Czernichow P, Levy-Marchal C. Insulin resistance early in adulthood in subjects born with intrauterine growth retardation. *J. Clin. Endocrinol. Metab.* 85(4), 1401–1406 (2000).
- 134 Holemans K, Aerts L, Van Assche FA. Lifetime consequences of abnormal fetal pancreatic development. *J. Physiol.* 547(1), 11–20 (2003).
- 135 Snoeck A, Remacle C, Reusens B, Hoet JJ. Effect of a low protein diet during pregnancy on the fetal rat endocrine pancreas. *Biol. Neonate* 57(2), 107–118 (1990).
- 136 Garofano A, Czernichow P, Bréant B. *In utero* undernutrition impairs rat β -cell development. *Diabetologia* 40(10), 1231–1234 (1997).
- 137 Garofano A, Czernichow P, Bréant B. β -cell mass and proliferation following late fetal and early postnatal malnutrition in the rat. *Diabetologia* 41(9), 1114–1120 (1998).
- 138 Fowden AL, Giussani DA, Forhead AJ. Endocrine and metabolic programming during intrauterine development. *Early Hum. Dev.* 81(9), 723–734 (2005).
- 139 Tarry-Adkins JL, Martin-Gronert MS, Chen JH, Cripps RL, Ozanne SE. Maternal diet influences DNA damage, aortic telomere length, oxidative stress, and antioxidant defense capacity in rats. *FASEB J.* 22(6), 2037–2044 (2008).
- 140 Park JH, Stoffers DA, Nicholls RD, Simmons RA. Development of Type 2 diabetes following intrauterine growth retardation in rats is associated with progressive epigenetic silencing of Pdx1. *J. Clin. Invest.* 118(6), 2316–2324 (2008).
- 141 Martin JF, Johnston CS, Han C-T, Benyshek DC. Nutritional origins of insulin resistance: a rat model for diabetes-prone human populations. *J. Nutr.* 130(4), 741–744 (2000).
- 142 Blondeau B, Avril I, Duchene B, Bréant B. Endocrine pancreas development is altered in foetuses from rats previously showing intra-uterine growth retardation in response to malnutrition. *Diabetologia* 45(3), 394–401 (2002).
- 143 Seckl JR. Glucocorticoids, developmental ‘programming’ and the risk of affective dysfunction. *Prog. Brain Res.* 167, 17–34 (2008).
- 144 Li J, Forhead AJ, Dauncey MJ, Gilmour RS, Fowden AL. Control of growth hormone receptor and insulin-like growth factor-I expression by cortisol in ovine fetal skeletal muscle. *J. Physiol.* 541(2), 581–589 (2002).
- 145 Li J, Gilmour RS, Saunders JC, Dauncey MJ, Fowden AL. Activation of the adult mode of ovine growth hormone receptor gene expression by cortisol during late fetal development. *FASEB J.* 13(3), 545–552 (1999).
- 146 Li J, Owens JA, Owens PC, Saunders JC, Fowden AL, Gilmour RS. The ontogeny of hepatic growth hormone receptor and insulin-like growth factor I gene expression in the sheep fetus during late gestation: developmental regulation by cortisol. *Endocrinology* 137(5), 1650–1657 (1996).
- 147 Seckl JR. Glucocorticoids, feto-placental 11β -hydroxysteroid dehydrogenase type 2, and the early life origins of adult disease. *Steroids* 62(1), 89–94 (1997).
- 148 Lingas R, Dean F, Matthews SG. Maternal nutrient restriction (48 h) modifies brain corticosteroid receptor expression and endocrine function in the fetal guinea pig. *Brain Res.* 846(2), 236–242 (1999).
- 149 Lesage J, Bondeau B, Grino M, Breant B, Dupouy JP. Maternal undernutrition during late gestation induces fetal overexposure to glucocorticoids and intrauterine growth retardation, and disturbs the hypothalamo-pituitary-adrenal axis in the newborn rat. *Endocrinology* 142, 1692–1702 (2001).
- 150 Jaquiere AL, Oliver MH, Bloomfield FH, Connor KL, Challis JRG, Harding JE. Fetal exposure to excess glucocorticoid is unlikely to explain the effects of periconceptual undernutrition in sheep. *J. Physiol.* 572(1), 109–118 (2006).
- 151 Seckl JR, Holmes MC. Mechanisms of disease: glucocorticoids, their placental metabolism and fetal ‘programming’ of adult pathophysiology. *Nat. Clin. Pract. End. Met.* 3(6), 479–488 (2007).
- 152 Connor KL, Challis JRG, van Zijl PL *et al.* Do alterations in placental 11β -hydroxysteroid dehydrogenase (11β -HSD) activities explain differences in

- fetal hypothalamic-pituitary-adrenal (HPA) function following periconceptual undernutrition or twin conception in sheep? *Reprod. Sci.* 16(12), 1201–1212 (2009).
- 153 Bloomfield FH, Harding JE. Evidence for fetal glucocorticoid excess as a cause of adult cardiovascular disease. *Curr. Opin. Endocrinol. Diabetes Care* 13(6), 523–529 (2006).
- 154 Jaquiere AL, Oliver MH, Rumball CWH, Bloomfield FH, Harding JE. Undernutrition before mating in ewes impairs the development of insulin resistance during pregnancy. *Obstet. Gynecol.* 114(4), 869–876 (2009).
- 155 Challis JR, Sloboda D, Matthews SG *et al.* The fetal-placental hypothalamic–pituitary–adrenal (HPA) axis, parturition and postnatal health. *Mol. Cell. Endocrinol.* 185(1–2), 135–144 (2001).
- 156 Lesage J, Sebaai N, Leonhardt M *et al.* Perinatal maternal undernutrition programs the offspring hypothalamo–pituitary–adrenal (HPA) axis. *Stress* 9(4), 183–198 (2006).
- 157 Dong F, Ford SP, Nijland MJ, Nathanielsz PW, Ren J. Influence of maternal undernutrition and overfeeding on cardiac ciliary neurotrophic factor receptor and ventricular size in fetal sheep. *J. Nutr. Biochem.* 19(6), 409–414 (2008).
- 158 Bloomfield FH, Oliver MH, Hawkins P *et al.* Periconceptual undernutrition in sheep accelerates maturation of the fetal hypothalamic–pituitary–adrenal axis in late gestation. *Endocrinology* 145(9), 4278–4285 (2004).
- 159 Connor KL, Bloomfield FH, Oliver MH, Harding JE, Challis JRG. Effect of periconceptual undernutrition in sheep on late gestation expression of mRNA and protein from genes involved in fetal adrenal steroidogenesis and placental prostaglandin production. *Reprod. Sci.* 16(6), 573–583 (2009).
- 160 Edwards LJ, Bryce AE, Coulter CL, McMillen IC. Maternal undernutrition throughout pregnancy increases adrenocorticotropic receptor and steroidogenic acute regulatory protein gene expression in the adrenal gland of twin fetal sheep during late gestation. *Mol. Cell. Endocrinol.* 196(1–2), 1–10 (2002).
- 161 Harris HJ, Kotelevtsev Y, Mullins JJ, Seckl JR, Holmes MC. Intracellular regeneration of glucocorticoids by 11 β -hydroxysteroid dehydrogenase (11 β -HSD)-I plays a key role in regulation of the hypothalamic–pituitary–adrenal axis: analysis of 11 β -HSD-I deficient mice. *Endocrinology* 142(1), 114–120 (2001).
- 162 Bloomfield FH, Phua HH, Connor KL *et al.* Twins of periconceptually undernourished (PCUN) ewes have delayed adrenal maturation throughout gestation and decreased hypothalamic–pituitary–adrenal axis (HPAA) negative feedback at the level of the hippocampus. *J. Paediatr. Child Health* 44 (Suppl. 1), A39 (2008).
- 163 Stevens A, Cook A, Rumball CWH *et al.* Periconceptual undernutrition is associated with epigenetic changes in the *POMC* gene in the hypothalamus of fetal sheep. *Endocrine Abstracts* 19, P141 (2009).
- 164 Stevens A, Cook A, Rumball CWH *et al.* Epigenetic changes of the hypothalamic glucocorticoid receptor gene occur in the ovine fetus in conjunction with periconceptual undernutrition. *Endocrine Abstracts* 19, P142 (2009).
- 165 Sebaai N, Lesage J, Breton C, Vieau D, Deloof S. Perinatal food deprivation induces marked alterations of the hypothalamo–pituitary–adrenal axis in 8-month-old male rats both under basal conditions and after a dehydration period. *Neuroendocrinology* 79(4), 163–173 (2004).
- 166 Dutriez-Casteloot I, Breton C, Coupe B *et al.* Tissue-specific programming expression of glucocorticoid receptors and 11 β -HSDs by maternal perinatal undernutrition in the HPA axis of adult male rats. *Horm. Metab. Res.* 40(4), 257–261 (2008).
- 167 Chadio SE, Kotsampasi B, Papadomichelakis G *et al.* Impact of maternal undernutrition on the hypothalamic–pituitary–adrenal axis responsiveness in sheep at different ages postnatal. *J. Endocrinol.* 192(3), 495–503 (2007).
- 168 Oliver M, Todd S, Bloomfield F, Jaquiere A, Harding J. Cortisol response to AVP+CRH challenge is reduced in lambs born to ewes undernourished during the periconceptual period. *J. Paediatr. Child Health* 44(Suppl. 1), A39 (2008).
- 169 Sloboda DM, Moss TJ, Gurrin LC, Newnham JP, Challis JR. The effect of prenatal betamethasone administration on postnatal ovine hypothalamic–pituitary–adrenal function. *J. Endocrinol.* 172(1), 71–81 (2002).
- 170 Sloboda DM, Moss TJ, Li S *et al.* Prenatal betamethasone exposure results in pituitary–adrenal hyporesponsiveness in adult sheep. *Am. J. Physiol. Endocrinol. Metab.* 292(1), E61–E70 (2007).
- 171 Bauer MK, Breier BH, Harding JE, Veldhuis JD, Gluckman PD. The fetal somatotrophic axis during long term maternal undernutrition in sheep: evidence for nutritional regulation *in utero*. *Endocrinology* 136(3), 1250–1257 (1995).
- 172 Waters MJ, Kaye PL. The role of growth hormone in fetal development. *Growth Horm. IGF Res.* 12(3), 137–146 (2002).
- 173 Woods K, Camacho-Hubner C, Savage MO, Clarke AJL. Intrauterine growth retardation and postnatal growth failure associated with deletion of the insulin-like growth factor 1 gene. *N. Engl. J. Med.* 335(18), 1363–1367 (1996).
- 174 Russell WE. Endocrine and other factors affecting growth. In: *Fetal and Neonatal Physiology*. Polin RA, Fox WW (Eds). WB Saunders Company, PA, USA 204–213 (1992).
- 175 D'Ercole AJ, Stiles AD, Underwood LE. Tissue concentrations of somatomedin C: further evidence for multiple sites of synthesis and paracrine or autocrine mechanisms of action. *Proc. Natl Acad. Sci. USA* 81(3), 935–939 (1984).
- 176 Nissley SP. Growth factors. In: *Principles and Practice of Endocrinology and Metabolism*. Becker KL, Bilezikian JP, Bremner WJ *et al.* (Eds.). JB Lippincott Company, PA, USA 1315–1321 (1990).
- 177 Baker J, Liu J-P, Robinson EJ, Efstratiadis A. Role of insulin-like growth factors in embryonic and postnatal growth. *Cell* 75, 79–82 (1993).
- 178 Bloomfield FH, Harding JE. Experimental aspects of nutrition and fetal growth. *Fet. Mat. Med. Rev.* 10(2), 91–107 (1998).
- 179 Liu J-P, Baker J, Perkins AS, Robinson EJ, Efstratiadis A. Mice carrying null mutations of the genes encoding insulin-like growth factor I (IGF-I) and type I IGF receptor (IGF1r). *Cell* 75, 59–72 (1993).
- 180 Fowden AL. The insulin-like growth factors and foeto–placental growth. *Placenta* 24(8–9), 803–812 (2003).
- 181 Osgerby JC, Wathes DC, Howard D, Gadd TS. The effect of maternal undernutrition on ovine fetal growth. *J. Endocrinol.* 173(1), 131–141 (2002).
- 182 Ward MA, Neville TL, Reed JJ *et al.* Effects of selenium supply and dietary restriction on maternal and fetal metabolic hormones in pregnant ewe lambs. *J. Anim. Sci.* 86(5), 1254–1262 (2008).
- 183 Oliver MH, Harding JE, Breier BH, Evans PC, Gluckman PD. Glucose but not a mixed amino acid infusion regulates

- plasma insulin-like growth factor-I concentrations in fetal sheep. *Pediatr. Res.* 34(1), 62–65 (1993).
- 184 Oliver MH, Harding JE, Breier BH, Gluckman PD. Fetal insulin-like growth factor (IGF)-I and IGF-II are regulated differently by glucose or insulin in the sheep fetus. *Reprod. Fertil. Dev.* 8(1), 167–172 (1996).
- 185 Liu L, Harding JE, Evans PC, Gluckman PD. Maternal insulin-like growth factor-I infusion alters fetoplacental carbohydrate and protein metabolism in pregnant sheep. *Endocrinology* 135(3), 895–900 (1994).
- 186 Holt RIG. Fetal programming of the growth hormone-insulin-like growth factor axis. *Trends Endocrinol. Metabol.* 13(9), 392–397 (2002).
- 187 Gallaher BW, Breier BH, Keven CL, Harding JE, Gluckman PD. Fetal programming of insulin-like growth factor (IGF)-I and IGF-binding protein-3: evidence for an altered response to undernutrition in late gestation following exposure to periconceptual undernutrition in the sheep. *J. Endocrinol.* 159, 501–508 (1998).
- 188 Woodall SM, Breier BH, Johnston BM, Gluckman PD. A model of intrauterine growth retardation caused by chronic maternal undernutrition in the rat: effects on the somatotrophic axis and postnatal growth. *J. Endocrinol.* 150(2), 231–242 (1996).
- 189 Rae MT, Rhind SM, Kyle CE, Miller DW, Brooks AN. Maternal undernutrition alters triiodothyronine concentrations and pituitary response to GnRH in fetal sheep. *J. Endocrinol.* 173(3), 449–455 (2002).
- 190 Gatford KL, Clarke IJ, De Blasio MJ, McMillen IC, J SR, Owens JA. Perinatal growth and plasma GH profiles in adolescent and adult sheep. *J. Endocrinol.* 173(1), 151–159 (2002).
- 191 Woods KA, Van Helvoirt M, Ong KKL *et al.* The somatotrophic axis in short children born small for gestational age: relation to insulin resistance. *Pediatr. Res.* 51(1), 76–80 (2002).
- 192 Chatelain PG, Nicolino M, Claris O, Salle B, Chaussain J. Multiple hormone resistance in short children born with intrauterine growth retardation? *Horm. Res.* 49(Suppl. 2), 20–22 (1998).
- 193 de Waal WJ, Hokken-Koelega ACS, Stijnen T, Muinck Keizer-Schrama SMPF, Dropt SLS. Endogenous and stimulated GH secretion, urinary GH excretion, and plasma IGF-I and IGF-II levels in prepubertal children with short stature after intrauterine growth retardation. *Clin. Endocrinol.* 41(5), 621–630 (1994).
- 194 Flanagan DE, Moore VM, Godsland IF, Cockington RA, Robinson JS, Phillips DIW. Reduced foetal growth and growth hormone secretion in adult life. *Clin. Endocrinol.* 50, 735–740 (1999).
- 195 Huizinga CT, Oudejans CB, Delemarre-van de Waal HA. Persistent changes in somatostatin and neuropeptide Y mRNA levels but not in growth hormone-releasing hormone mRNA levels in adult rats after intrauterine growth retardation. *J. Endocrinol.* 168(2), 273–281 (2001).
- 196 Huizinga CT, Oudejans CBM, Steiner RA, Clifton ADK, Delemarre-Van De Waal HA. Effects of intrauterine and early postnatal growth restriction on hypothalamic somatostatin gene expression in the rat. *Pediatr. Res.* 48(6), 815–820 (2000).
- 197 Huizinga CT, Oudejans CB, Delemarre-Van de Waal HA. Decreased galanin mRNA levels in growth hormone-releasing hormone neurons after perinatally induced growth retardation. *J. Endocrinol.* 170(3), 521–528 (2001).
- 198 Burrow GN, Fisher DA, Larsen PR. Maternal and fetal thyroid function. *N. Engl. J. Med.* 331(16), 1072–1078 (1994).
- 199 Chan S, Kilby MD. Thyroid hormone and central nervous system development. *J. Endocrinol.* 165(1), 1–8 (2000).
- 200 Symonds ME. Pregnancy, parturition and neonatal development: interactions between nutrition and thyroid hormones. *Proc. Nutr. Soc.* 54(2), 329–343 (1995).
- 201 Dwyer CM, Stickland NC. The effects of maternal undernutrition on maternal and fetal serum insulin-like growth factors, thyroid hormones and cortisol in the guinea pig. *J. Dev. Physiol.* 18(6), 303–313 (1992).
- 202 De Blasio MJ, Gatford KL, Robinson JS, Owens JA. Placental restriction alters circulating thyroid hormone in the young lamb postnatally. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 291(4), R1016–R1024 (2006).
- 203 Chan SY, Andrews MH, Lingas R *et al.* Maternal nutrient deprivation induces sex-specific changes in thyroid hormone receptor and deiodinase expression in the fetal guinea pig brain. *J. Physiol.* 566(2), 467–480 (2005).
- 204 Thorpe-Beeston JG, Nicolaides KH, Snijders RJ, Felton CV, McGregor AM. Thyroid function in small for gestational age fetuses. *Obstet. Gynecol.* 77(5), 701–706 (1991).
- 205 Kilby MD, Verhaeg J, Gittoes N, Somerset DA, Clark PMS, Franklyn JA. Circulating thyroid hormone concentrations and placental thyroid hormone receptor expression in normal human pregnancy and pregnancy complicated by intrauterine growth restriction (IUGR). *J. Clin. Endocrinol. Metab.* 83(8), 2964–2971 (1998).
- 206 Kilby MD, Gittoes N, McCabe C, Verhaeg J, Franklyn JA. Expression of thyroid receptor isoforms in the human fetal central nervous system and the effects of intrauterine growth restriction. *Clin. Endocrinol.* 53(4), 469–477 (2000).
- 207 Mahajan SD, Aalinkel R, Singh S, Shah P, Gupta N, Kochupillai N. Thyroid hormone dysregulation in intrauterine growth retardation associated with maternal malnutrition and/or anemia. *Horm. Metab. Res.* 37(10), 633–640 (2005).
- 208 Jacobsen BB, Hummer L. Changes in serum concentrations of thyroid hormones and thyroid hormone-binding proteins during early infancy: studies in healthy fullterm, small-for-gestational age and preterm infants aged 7 to 240 days. *Acta Paediatr.* 68(4), 411–418 (1979).
- 209 Jacobsen BB, Andersen HJ, Peitersen B, Dige-Petersen H, Hummer L. Serum levels of thyrotropin, thyroxine and triiodothyronine in fullterm, small-for-gestational age and preterm newborn babies. *Acta Paediatr.* 66(6), 681–687 (1977).
- 210 Fetoui H, Bouaziz H, Mahjoubi-Samet A, Soussia L, Guermazi F, Zeghal N. Food restriction induced thyroid changes and their reversal after refeeding in female rats and their pups. *Acta Biol. Hung.* 57(4), 391–402 (2006).
- 211 Ramos CF, Lima APS, Teixeira CV, Brito PD, Moura EG. Thyroid function in post-weaning rats whose dams were fed a low-protein diet during suckling. *Braz. J. Med. Biol. Res.* 30, 133–137 (1997).
- 212 Passos MCF, da Fonte Ramos C, Dutra SCP, Mouço T, de Moura EG. Long-term effects of malnutrition during lactation on the thyroid function of offspring. *Horm. Metab. Res.* 34(1), 40–43 (2002).
- 213 Moura E, Passos M. Neonatal programming of body weight regulation and energetic metabolism. *Biosci. Rep.* 25, 251–269 (2005).

- 214 de Escobar GM, Obregón MJ, del Rey FE. Iodine deficiency and brain development in the first half of pregnancy. *Public Health Nutr.* 10(12A), 1554–1570 (2007).
- 215 Boyages SC, Halpern J-P, Maberly GF *et al.* A comparative study of neurological and myxedematous endemic cretinism in western China. *J. Clin. Endocrinol. Metab.* 67(6), 1262–1271 (1988).
- 216 Black RE, Allen LH, Bhutta ZA *et al.* Maternal and child undernutrition: global and regional exposures and health consequences. *Lancet* 371(9608), 243–260 (2008).
- 217 Inskip HM, Crozier SR, Godfrey KM *et al.* Women's compliance with nutrition and lifestyle recommendations before pregnancy: general population cohort study. *BMJ* 338(7694), 586–589 (2009).
- 218 Lillycrop KA, Phillips ES, Jackson AA, Hanson MA, Burdge GC. Dietary protein restriction of pregnant rats induces and folic acid supplementation prevents epigenetic modification of hepatic gene expression in the offspring. *J. Nutr.* 135(6), 1382–1386 (2005).

Affiliations

- Jane E Harding
Liggins Institute, University of Auckland,
Private Bag 92019, Auckland,
New Zealand
Tel.: +64 9373 7599 ext 86439
Fax: +64 9 373 7497
j.harding@auckland.ac.nz
- José GB Derraik
Liggins Institute, University of Auckland,
Private Bag 92019, Auckland,
New Zealand
derraik@gmail.com
- Frank H Bloomfield
Liggins Institute, University of Auckland,
Private Bag 92019, Auckland,
New Zealand
Tel.: +64 9 373 7599 ext 86107
Fax: +64 9 373 8763
f.bloomfield@auckland.ac.nz